

# $^{17}\text{O}$ imaging for the evaluation of physiological function in the tumor bearing mice

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**Introduction:** The application of  $^{17}\text{O}$  MRI method has been proposed in the measurements of regional cerebral blood flow and cerebral metabolic rate of oxygen consumption<sup>1-3</sup>. Since hypoxic tumor cells are resistant to the radiation, monitoring the tumor oxygenation and blood flow is important for the effective treatment. Irrespective of such expectation, low sensitivity and short  $T_2$  of  $^{17}\text{O}$  make the direct measurement difficult. In order to overcome these difficulties, proton-detected  $^{17}\text{O}$  MRI<sup>4</sup>, Chemical Shift Imaging<sup>5</sup>, gradient echo and projection reconstruction<sup>6</sup> have been proposed. In this study, FISP was employed for  $\text{H}_2^{17}\text{O}$  imaging at 7.0 T.

**Methods:** A water phantom (70 ml, natural abundance  $^{17}\text{O}$ ) was used for sequence comparison, FISP, Flash and FSE, and optimization. Saline of 0.5 ml containing 1 – 5%  $^{17}\text{O}$  prepared from 10%  $^{17}\text{O}$  water (Cambridge Isotope Laboratories, Inc) was i.v. injected to tumor bearing C3H/He mice (25 – 30 g). MRS/MRI was performed under ketamine and xylazine anesthesia on 7T/400mm/SS system (NIRS/KOBELCO/Bruker) with 50 mm  $^1\text{H}/^{17}\text{O}$  Litz coil (Doty Scientific Inc.).  $^{17}\text{O}$  images of mice were obtained by true FISP under the following parameters; TR/TE 6.54/3.28 ms, flip angle 60°, FOV 32×8 cm, matrix size 128×32, 100 or 15 mm slice thickness. After imaging experiment, organs were excised for  $^{17}\text{O}$  spectral measurements.

**Results:** Estimated  $T_2^*$  from FWHM of  $\text{H}_2^{17}\text{O}$  in a phantom and a mouse were 6 and 2 ms, respectively. The S/N in the image of a water phantom by FISP was 1.6 times higher than that by Flash (TR/TE 10.1/5.2 ms and flip angle 75°) for the same data acquisition time, and the minimum TE was still too long for  $\text{H}_2^{17}\text{O}$  by FSE. FISP  $^{17}\text{O}$ -Image of a tumor bearing mouse with natural abundance  $\text{H}_2^{17}\text{O}$  in 10 min is shown in Fig.1a. After the injection of 5%  $^{17}\text{O}$  saline, the whole body signal intensities of both  $^{17}\text{O}$  spectra and images were increased ca. 10 times higher than pre injection, which was sufficient for the visualization of the organ profile by  $^{17}\text{O}$  images (Fig. 1b). Whole body image intensity was still more than 50% of maximum value 15 hours after the injection (Fig.1c). The signal intensities of tumor in *in vivo* image and *ex vivo* spectrum at 15 h after injection were 6 and 4.5 times higher than those from non- $^{17}\text{O}$  saline injected mouse, respectively.

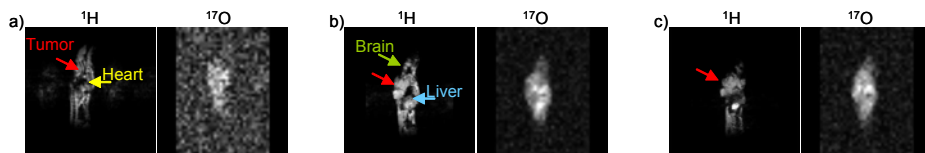


Fig.1.  $^{17}\text{O}$  images (sagittal) of a tumor bearing mouse on the back at pre (a), 15 min (b) and 15 h (c) after 5%  $^{17}\text{O}$  saline injection, without window functions.

**Discussion:** FISP was the best in imaging of  $^{17}\text{O}$  water among FISP, Flash, and FSE. The low resonance frequency of  $^{17}\text{O}$ , 40 MHz at 7 T, brought us the advantage of high sensitivity of FISP over the susceptibility problem. The distribution of  $\text{H}_2^{17}\text{O}$  in mouse was successfully visualized by FISP in 10 min under a spatial resolution of 2.5 mm. The regional  $^{17}\text{O}$  image intensities were in good agreement with the  $^{17}\text{O}$  NMR spectral intensities of excised organs. The increases in intensity both in the images and spectra after  $^{17}\text{O}$  saline injection were in proportional to the increase in the  $^{17}\text{O}$  concentration in the extracellular fluid. The feature of  $^{17}\text{O}$  signals observed here reports the physical state of water *in vivo*, which is to be studied further. Here, a feasibility of  $\text{H}_2^{17}\text{O}$  mapping depicting water dynamics from blood flow to diffusion is shown. The extension of this technique to visualize the oxygen consumption in the brain and tumor using  $^{17}\text{O}_2$  gas is promising.

**Reference:** (1) Zhu XH. et al, NMR in Biomed 18:83 (2005), (2) Ronen I. et al, PNAS 95:12934 (1998), (3) Arai T. et al, Brain Res 45:451 (1998), (4) Ronen I. et al. MRM 32:789 (1994), (5) Xhu XH. et al, MRM 45:543 (2001), (6) Fiat D. et al, Neurol Res 26:803 (2004)