

In Vivo Measurement of the Longitudinal Relaxation Time of Arterial Blood (T_{1a}) in the Mouse using a Pulsed Arterial Spin Labeling Approach

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INTRODUCTION

The longitudinal relaxation time of arterial blood water (T_{1a}) is a parameter that is required for the quantification of cerebral blood flow (CBF) using arterial spin labelling (ASL) techniques. However, T_{1a} is difficult to measure *in vivo*, due to the high speed of blood flow through the arteries and the small size of the arteries (particularly in small animals) which causes partial volume effects within voxels. Further, *ex vivo* measurements do not exactly replicate the *in vivo* conditions, and therefore may provide incorrect results. In this work, we have developed and applied a method based on pulsed ASL for measuring T_{1a} *in vivo*. The advantage of this approach is that it uses differences in tissue magnetisation to estimate T_{1a} , thereby eliminating the requirement to image and isolate intravascular blood signal.

METHODS

The FAIR technique with a global pre-saturation pulse was used, as has been described previously (1). With this sequence, assuming all non-selective saturation and inversion pulses are truly global, the FAIR difference magnetisation ΔM is described by:

$$\Delta M(TI, \tau) = \frac{2M_0\alpha_0}{\left(\frac{1}{T_{1app}} - \frac{1}{T_{1a}}\right)} \frac{f}{\lambda} e^{-\delta/T_{1a}} \left(e^{-(TI-\delta)/T_{1a}} - e^{-(TI-\delta)/T_{1app}} \right) \left[1 - e^{-\tau/T_{1a}} \right] \quad \{1\}$$

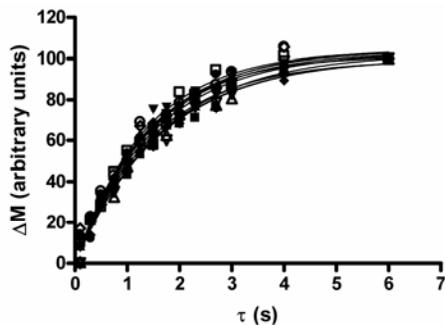
where TI is the inversion time, M_0 is spin density, α_0 is the inversion efficiency, f is CBF, λ is the blood:brain partition coefficient, δ is the transit time, T_{1app} is the apparent tissue T_1 following slice-selective inversion and

τ is the time delay between the global pre-saturation and the subsequent inversion pulse. It can be seen from the last term of equation {1} that if measurements of ΔM are made over a range of τ , the data should follow a simple monoexponential recovery with T_{1a} as the time constant.

MRI studies were performed on anaesthetised adult male CD1 mice (n=5, 30-35g) using a 2.35T horizontal magnet interfaced to a SMIS console. RF excitation pulses were transmitted via a volume coil (6cm length) and received by a separate passively decoupled single loop surface coil (15mm diameter). FAIR images were acquired using single-shot spin echo EPI. A train of 4 adiabatic half sech pulses, each followed by a spoiler gradient, was used for global pre-saturation. FOCI inversion pulses (2) were used for efficient spin labelling and optimal slice definition. FAIR image pairs were acquired with a TI of 1.3s and the following values of τ (s): 0.1, 0.3, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 2.3, 2.7, 3, 4, and 6. The acquisition order was randomized and different for each animal. Imaging parameters were: slice thickness = 2mm; image matrix size = 128x64; field of view = 32x16mm²; TE = 36ms; N_{av} = 32. The total acquisition time for all τ values was approximately 55 minutes. A bipolar gradient pulse ($b \approx 17s/mm^2$) was used to eliminate intravascular signal from large blood vessels. Analysis was performed on ROIs placed in cortical grey matter regions.

RESULTS

The FAIR subtraction data $\Delta M(\tau)$ and fits to the theoretical model are shown in the figure below (left). It is clear that the variation of ΔM with τ is well described by a monoexponential recovery. The T_{1a} values resulting from the fits for all the animals are shown in the Table below. The average value of T_{1a} from the fits was **1.51 ± 0.11s** (mean ± SD). Also shown in the table are the corresponding brain tissue T_1 values for each mouse. The average value of brain tissue T_1 was **0.92 ± 0.04s** (mean ± SD), which is significantly less than the measured value of T_{1a} ($p < 0.0001$). This demonstrates that the recovery rate of ΔM as a function of τ reflects something different from tissue T_1 and is consistent with previous measurements of arterial blood T_1 .



Animal no.	ROI in cortex	
	T_{1a} (s)	Tissue T_1 (s)
1	1.58	0.90
2	1.51	0.90
3	1.47	0.88
4	1.60	0.99
5	1.40	0.92
Mean ± SD	1.51 ± 0.11	0.92 ± 0.04

DISCUSSION AND CONCLUSION

A new method for the *in vivo* measurement of arterial blood T_1 has been proposed and applied in the mouse. The method uses the tissue FAIR signal difference, and therefore does not require direct measurement of the intravascular blood signal. As a result, many of the problems associated with T_{1a} measurements are avoided. Consequently, T_{1a} can be measured directly (rather than extrapolated from literature values obtained at different field strengths), improving the reliability and accuracy of CBF quantification using ASL.

REFERENCES

- (1) Pell, Thomas, Lythgoe *et al.* MRM **41**, 829-40 (1999) (2) Ordidge, Wylezinska, Hugg *et al.* MRM **36**, 562-6 (1996)