

Doping Gd-DTPA Solution with Water-Soluble Fullerene C₈₂(OH)_n Increases the T₁ Relaxation Rate of Water Proton: In vitro and In vivo Studies

X. X. Wang¹, G. M. Xing², B. Zhang², Y. L. Zhao², H. Lei¹

¹State Key Laboratory of Magnetic Resonance and Atomic and Molecular Physics, Wuhan Institute of Physics & Mathematics, The Chinese Academy of Science, Wuhan, Hubei, China, People's Republic of, ²Institute of High Energy Physics, The Chinese Academy of Science, Beijing, China, People's Republic of

Introduction

With the advent of gadolinium (Gd)-based MRI contrast agents, many efforts have been given for the development of new complex compounds of Gd, such as water-soluble metallofullerenes Gd@C₈₂(OH)₄₀ and Gd@C₆₀[C(COOH)₂]₁₀ [1,2]. It has been shown that metallofullerenes often possess much higher relaxivity than the commonly used Gd-chelate complexes [1,2], an attribute that may be associated with the unique physical/chemical properties of fullerene. In this study, we investigated the effect of water-soluble empty fullerene cage C₈₂(OH)_n on the T₁ relaxation rate of water proton in Gd-DTPA solution by both in vitro and in vivo measurements.

Materials and Methods

All MR measurements were carried out a 4.7T/30cm Bruker Biospec scanner equipped with actively shielded gradients. A 75mm-diameter Bruker volume coil was used for both excitation and reception.

In vitro study: Commercially available Gd-DTPA solution (i.e., Magnevist) was mixed with custom-synthesized C₈₂(OH)_n (n≈36) stock solution and diluted by saline. The concentration of Gd-DTPA in all samples was kept constant at 0.1mM, while that of C₈₂(OH)_n varied at 0, 0.05, 0.1, and 0.15mM. T₁ measurement was performed on bundled sample-filled 5mm NMR tubes using an inversion-recovery spin echo imaging sequence with FOV 3×3 cm², matrix size 64×64, TR/TE 10s/12ms, slice thickness 2mm and 9 T1 values ranging from 15ms to 9s.

In vivo study: Six male Wistar rats (150-180g) were used. Each rat was anesthetized by i.p. injection of chloral hydrate (20%, 0.3ml/100g body weight), and catheterized in the right femoral vein for contrast agent injection. Abdominal T₁-weighted spin echo images were acquired continuously before and after intravenous injection of either 0.75ml of 1mmol/L Gd-DTPA solution (N=3) or mixed solution 0.75ml of 1mmol/L Gd-DTPA and 0.25ml of 0.4mmol/L C₈₂(OH)_n (N=3). The imaging parameters were: FOV 5×5cm², matrix size 128×128, TR/TE 300ms/14ms, slice thickness 1.5mm and 5 averages.

Results

The T₁ relaxation rate of water proton (R₁) in the mixed solution of Gd-DTPA and C₈₂(OH)_n as a function of C₈₂(OH)_n concentration is shown in Fig. 1. Figure 2 shows the T₁-weighted abdominal images of two rats before and 7 minutes after injection of contrast agents. In the rat injected with Gd-DTPA only (Fig. 2, rat1), the signal enhancements observed for renal cortex, renal calyces and liver were 9.6%, 46.4% and 2.4%, respectively. In the rat injected with Gd-DTPA/C₈₂(OH)_n mixture (Fig. 2, rat2), the signal intensities of renal cortex, renal calyces and liver increased 37.9%, 163.3% and 4.8%, respectively. Half hour after injections, the signal enhancements in renal cortex, renal calyces and liver were 1.7%, 27.8 % and -1.6% for the Gd-DTPA injected rat, respectively, and 35.3%, 216.5% and 3.2% for the Gd-DTPA/C₈₂(OH)_n mixture injected rat.

Discussion

The results of the phantom experiments show that T₁ relaxation rate of water proton in C₈₂(OH)_n-doped Gd-DTPA solution increases linearly with the concentration of C₈₂(OH)_n. A putative non-covalent interaction between Gd-DTPA and C₈₂(OH)_n, which slows down the rotational correlation time of either Gd-DTPA or water, or both, and leads to increased exchange rate, might underlie the observation. Compared to the rats injected with the same dose of Gd-DTPA alone, the rats injected with Gd-DTPA/C₈₂(OH)_n mixture showed stronger signal intensity enhancement in both kidney and liver (Fig. 2), especially in the renal calyces where the signal enhancement was almost 4 times of that with Gd-DTPA injection. Signal enhancement caused by Gd-DTPA/C₈₂(OH)_n mixture injection also lasted much longer than that caused by Gd-DTPA injection, indicating a longer blood retention time and altered biodistribution of Gd-DTPA when interacted with C₈₂(OH)_n. The nature of the interaction between Gd-DTPA and polyhydroxyl fullerene needs further investigation.

Acknowledgement: Supported by Natural Science Foundation of China 10234070, 30370419 and 30400136.

References: 1) Mikawa M. et al, *Bioconjugate Chem.*, 12:510-514 (2001). 2) Bolskar RD. et al, *J. Am. Chem. Soc.*, 125:5471-5478 (2003).

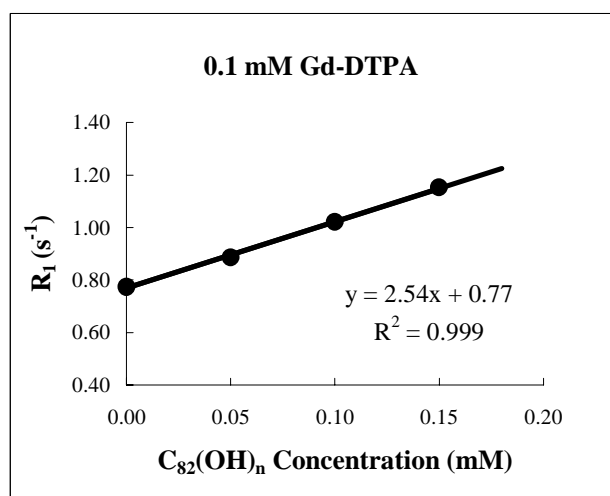


Figure 1. T₁ relaxation rate of water proton (R₁) in the mixed solution of Gd-DTPA and C₈₂(OH)_n has a linear relationship with the C₈₂(OH)_n concentration.

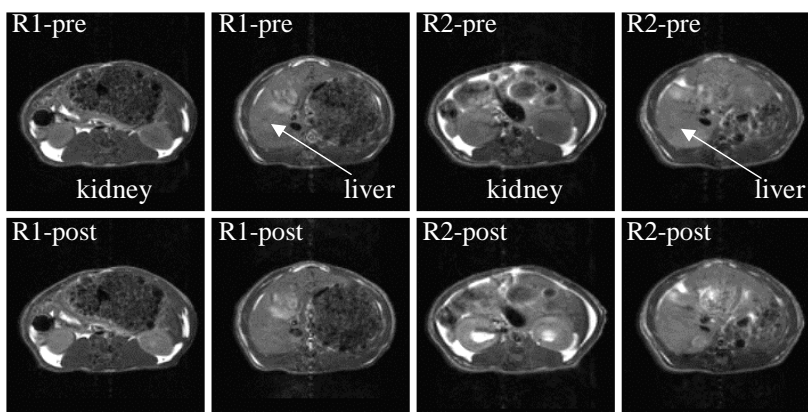


Figure 2. T₁-weighted abdominal images of two rats (R1 and R2) showing the kidneys and the liver before (-pre) and 7 minutes after (-post) intravenous injection of contrast agents. R1: 0.75ml of 1mmol/L Gd-DTPA; R2: 0.75ml of 1mmol/L Gd-DTPA mixed with 0.25ml of 0.4mmol/L C₈₂(OH)_n.