

# Estimating the time-course of brain adaptation and angiogenesis using PtO<sub>2</sub> measurements—an in vivo EPR study

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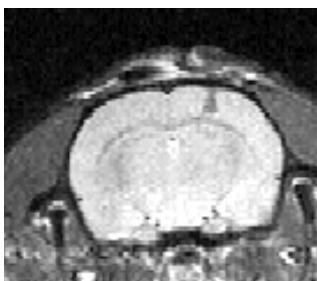
## INTRODUCTION

Chronic hypoxic exposure initiates a range of adaptive processes, including angiogenesis, upregulation of glycolysis and increased hematocrit [1]. There are no major changes in CMRO<sub>2</sub>. Since PtO<sub>2</sub> is determined by a balance between oxygen delivery and utilization, and utilization remains constant, then PtO<sub>2</sub> can be used to monitor adaptation. The time-course of brain PtO<sub>2</sub> during chronic and acute exposure to systemic hypoxia was investigated. A unique feature of this study, is that we can monitor brain PtO<sub>2</sub> in awake restrained animals using electron paramagnetic resonance [2].

## METHODS

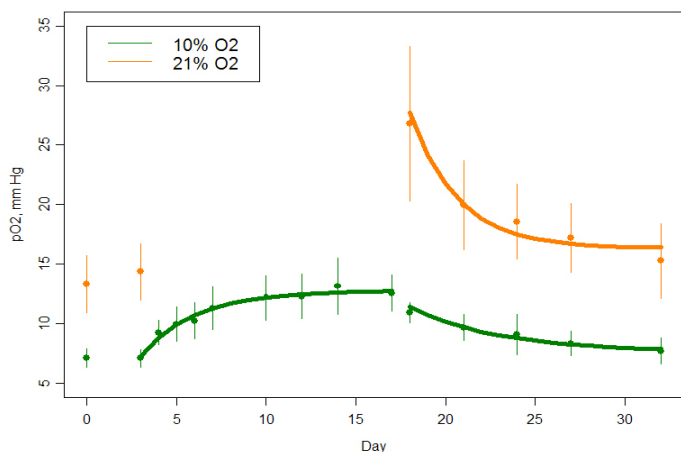
EPR measurements were done while animals (n=7, Wistar rats) breathed both 10% and 21% O<sub>2</sub>. Measurements were taken over a time-course of acclimation to chronic hypoxia (1/2 atm) and recovery. EPR methods have been previously described [2], and involve implantation of lithium phthalocyanine (LiPc) into the cortex, which has an EPR linewidth that is linearly proportional to pO<sub>2</sub> over 0 to 713 mmHg. EPR oximetry was conducted using a low frequency (1.18GHz, L-band) spectrometer with a surface-loop resonator. Awake restrained animals had PtO<sub>2</sub> measured while breathing 21% and 10% O<sub>2</sub> before and after acclimation. During acclimation, they were exposed to 10% FiO<sub>2</sub>. Acclimation was at a pressure of 370 mmHg, corresponding to a normobaric O<sub>2</sub> of 10%. Analysis of the data on the time-course of PtO<sub>2</sub> was accomplished using a nonlinear, exponential mixed effects model [3]. MRI for LiPc localization was done before the study began to confirm the material was within the cortex using T2w MRI, Varian console and a 7T Magnex magnet.

## RESULTS



MRI confirmed that LiPc was within the cortex (Fig. 1). The mean PtO<sub>2</sub> before acclimation breathing 21% O<sub>2</sub> was 13±2 and while breathing 10% O<sub>2</sub> was 7±2 mmHg (mean±SD, n=7). During acclimation, the PtO<sub>2</sub> was measured while breathing 10% O<sub>2</sub> to determine the time needed for brain PtO<sub>2</sub> to stabilize to a new level. The time required for the PtO<sub>2</sub> to reach a level not significantly different from control was 5-7 days with a T<sub>1/2</sub> for adaptation of 3.7 days. The time required for the PtO<sub>2</sub> to stabilize during deacclimation was 7-10 days, with a T<sub>1/2</sub> for deacclimation of 1.8 days (Fig. 2).

**Figure 1.** MRI of the LiPc implant showing cortical localization



**Figure 2.** Cortical PtO<sub>2</sub> measured while breathing 21% O<sub>2</sub> (top traces) and 10% O<sub>2</sub> (bottom traces) before acclimation (2 points), during acclimation (10% only) and during recovery. The curves represent the results of non-linear mixed effects statistical modeling (mean±SD, n=7)

## DISCUSSION

The PtO<sub>2</sub> represents the sum of many complex physiological processes. However, since CMRO<sub>2</sub> remains relatively constant, then it can be used to determine the sum of all adaptive processes during exposure to hypoxia. These data indicate that 5-7 days is sufficient for most adaptive processes during exposure, and that de-acclimation is also very rapid.

## REFERENCES

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2. Dunn, J.F., et al., *Non-invasive assessment of cerebral oxygenation during acclimation to hypobaric hypoxia*. J Cereb Blood Flow Metab, 2000. **20**: p. 1632-1635.
3. Demidenko, E., *Mixed Models: Theory and Application*. New York: Wiley, 2004.