

Mn²⁺-Enhanced ¹H₂O MRI Measurement of Na⁺/Ca²⁺-Exchanger Mediated Mn²⁺ Influx in the Rat Heart During Myocardial Hypoxia

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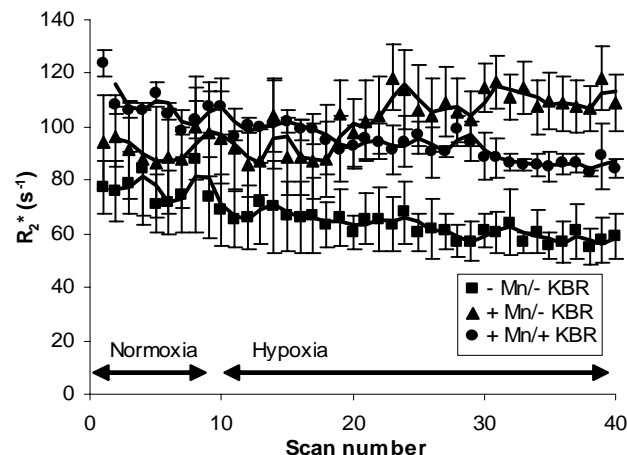
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Introduction: Data presented herein support the hypothesis that increases in free intra-cellular (i.c.) calcium, Ca²⁺, during hypoxia are mediated by the Na⁺/Ca²⁺-exchanger (NCX), which can be blocked with KBR-7943 (KBR), an NCX blocker specific for the reverse-direction¹. Hypoxia diminishes oxidative metabolism and lowers the i.c. pH as a result of extensive glycolysis. In addition, the hypoxia-induced cellular ATP deficit reduces Na⁺/K⁺-ATPase activity. In a compensatory attempt to restore cellular homeostasis, myocytes exchange i.c. H⁺ for extracellular (e.c.) Na⁺, via the Na⁺/H⁺-exchanger. The NCX, operating in “reverse” of the physiological normal net direction, exchanges i.c. Na⁺ for e.c. Ca²⁺ leading to a rise in free [Ca²⁺]_{i.c.}. The transmembrane Ca²⁺ influx can be traced with a true Ca²⁺ competitor², Mn²⁺, which is observed via its effect on the ¹H₂O MRI signal³. The myocardial ¹H₂O R₂ (≡ T₂⁻¹) value increases during hypoxia as long as the perfusion flow, and hence the [Mn²⁺]_{e.c.}, is maintained constant—thus allowing for myocardial Mn²⁺ uptake.

Methods: The apparent rate constant, R₂^{*}, is a measure of hyperfine plus susceptibility contributions to transverse relaxation. Values were measured for 9 rat hearts via ¹H₂O MRI (AMX 9.4 T Bruker, vertical bore system), before (10 scans) and after (30 scans) the onset of hypoxia, such that each heart served as its own control. The 9 rats were divided into 3 groups of 3 rats each: -Mn²⁺/-KBR, +Mn²⁺/-KBR, and +Mn²⁺/+KBR. The data were acquired with a single 1 mm thick transverse slice using a RARE sequence, α = 90°, TR = 12.3 s, TE = 0.023 s, FOV = (2 cm)², and matrix = (128)². Spin-density (S₀) and R₂^{*} values were obtained using the Paravision® mono-exponential fitting routine. Hearts were Langendorff-perfused^{4,5} with a 23°C modified Krebs-Henseleit solution—PO₄ and SO₄ were replaced with Cl⁻, and [K⁺]_{e.c.} was held at 15.75 mM to arrest the heart. The perfusion pressure was monitored constantly throughout each imaging session. The perfusing medium contained 25 μM Mn²⁺ whenever applicable.

Results: One of the rats was excluded from the group receiving KBR due to an incorrect Mn²⁺ loading of the heart. The mean normalized S₀ values among the 3 groups were not significantly different before or after hypoxia, according to the non-parametric Kruskal-Wallis test for k-independent samples (asympt. p-value = 0.875). The plot below depicts the mean R₂^{*} time-course for each group, before (scans 1 to 10) and after (scans 11 to 40) the onset of hypoxia. The mean R₂^{*} values (in s⁻¹), before and after hypoxia, for each group are: 76.5 (±6.0) and 62.4 (±4.9); 92.7 (±4.7) and 103.9 (±9.9); and 107.9 (±6.7) and 92.6 (±6.1), for the -Mn²⁺/-KBR (■), +Mn²⁺/-KBR (▲), and +Mn²⁺/+KBR (●) groups, respectively. After hypoxia, the mean R₂^{*} values for the control group (■) and that (●) receiving KBR are reduced by 18.4% and 14.2% relative to their respective baseline values. The mean R₂^{*} value for the group (▲) receiving Mn²⁺ but not KBR increased 12.1%, reflecting myocardial Mn²⁺ uptake. These differences are significant to p-value <0.001 according to Tukey and Bonferroni *Post Hoc* tests.

Conclusion: Decrements in control (■) and KBR-loaded (●) mean R₂^{*} values during the experiment reflect cardiac arrest/hypoxia-induced edema. Although S₀ values are expected to increase with edema, these changes were not significant according to the non-parametric Kruskal-Wallis test for k-independent samples. On the other hand, the mean R₂^{*} value for the group (▲) receiving Mn²⁺ but not KBR increases under hypoxia presumably because the rise in free paramagnetic [Mn²⁺]_{i.c.} catalyzes the loss of NMR coherence among ¹H₂O spins with which Mn²⁺ interacts, thus offsetting the edematous effects. The ability of Mn²⁺ to catalyze ¹H₂O R₂^{*} is strongly dependent on the reorientation time (τ_r) of its complex. For instance, protein-bound Mn²⁺ has a greater relaxivity than the Mn²⁺ aqua ion, because the former has a larger τ_r value.³ One may expect that increases in free [Ca²⁺]_{i.c.} and in its tracer Mn²⁺, would translate into greater or smaller R₂^{*} values depending on whether this rise stems from a Ca²⁺ influx or its release from intracellular populations, respectively. Therefore, the results reported herein are consistent with the hypothesis that increases in [Ca²⁺]_{i.c.} during hypoxia result from Ca²⁺ influx rather than its release from intracellular protein. [Though unlikely, R₂^{*} value reduction could be



accounted for by changes in the heart geometry and increased bulk magnetic susceptibility contributions to R₂^{*} during the experiments.] Since the nifedipine-sensitive slow Ca²⁺-channel^{2,6} is inoperative under hyperkalemic conditions and receptor mediated endocytosis of divalent metal cations⁷⁻⁹ is a slow process, Mn²⁺ enters the cell likely through the NCX during hypoxia—this is consistent with the observed inhibition of the Mn²⁺ effect on R₂^{*} by KBR.

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