

## Diffusion Tensor Imaging and T2 Relaxography in Myelin Deficient Mice

J. Nierenberg<sup>1,2</sup>, D. N. Guilfoyle<sup>1</sup>, M. J. Hoptman<sup>2,3</sup>, V. Dyakin<sup>1</sup>, R. A. Nixon<sup>2,4</sup>

<sup>1</sup>Medical Physics, Nathan Kline Institute, Orangeburg, NY, United States, <sup>2</sup>Psychiatry, NYU School of Medicine, New York, NY, United States, <sup>3</sup>Clinical Research, Nathan Kline Institute, Orangeburg, NY, United States, <sup>4</sup>Dementia Research, Nathan Kline Institute, Orangeburg, NY, United States

**INTRODUCTION-** Myelin abnormalities are present in a number of neurological diseases and may contribute to the pathophysiology of schizophrenia and Alzheimer's disease. Mouse models, such as the myelin basic protein (MBP) knockout *shiverer*, have been studied with MRI and neurohistology and represent an opportunity to better understand the impact of myelin-related defects on brain structure. Song et al. (Neuroimage, 17: 1429-36, 2002) used diffusion tensor imaging (DTI) in *shiverer in vivo* and showed that the MBP null mutation is associated with decreased diffusion anisotropy attributable to increased radial diffusivity. Here we extend these findings to *shiverer* fixed brain and report associations between MBP, DTI and T<sub>2</sub> relaxation measures.

**METHODS-** All mice used were 3-4 months of age. Following deep anesthesia with isoflurane, *shiverer* mice and their age-matched littermates (Jackson Labs) were perfused with 4% formalin-PBS. The top part of the skull was removed so that the head could be positioned and immobilized in a 15 ml centrifuge tube, filled with 4% formalin, such that the tube could be easily positioned at the coil's center. Postfixation continued for a minimum of 3 months. All the data were taken on a 7.0 T MRMS (Guildford, UK, formerly SMIS) 40 cm bore system. The imaging protocol and hardware details have been described in detail in Guilfoyle et al. NMR Biomed (2003) 16: 77-81. A 3-plane localizer was used initially to help standardize slice selection across samples and reliability was achieved. Briefly, coronal DTI scans (beginning at the callosal genu) were acquired using a distortion-free spin echo sequence (TR/TE 1000/40 ms, 128 x 128 matrix,  $b=900\text{s/mm}^2$ , 12 slices, 0.3mm slice thickness, 0mm gap). In-plane resolution was 85 $\mu\text{m}$ . Diffusion was measured along six equally spaced nonlinear directions and also with no diffusion gradients ( $b=0$  images). A multi-echo spin echo sequence was used for the T<sub>2</sub> measurements with a total of six separate echoes (in msec: 15, 20, 25, 30, 35, 55, 75) for the T<sub>2</sub> fit. Matrix size was 128 x 96 (zero-filled to 128 x 128) and TR was 2 s. The FOV, slice thickness and slice selection of the T<sub>2</sub> measurements were the same as used in the DTI experiments. ROIs were created on  $b=0$  images as they retained sufficient white matter contrast to define regional boundaries (though this was clearly diminished in *shiverer*). ROIs for white matter regions- corpus callosum (CC), anterior commissure (AC), internal capsule (IC), optic tract (OT) and fornix (FX)- and cerebral cortex (as a negative gray matter control, CTX) were then transferred to DTI-derived maps of fractional anisotropy (FA), trace diffusivity, axial (Dax) and radial  $[(\lambda_2+\lambda_3)/2]$  diffusivity (DRa) bilaterally. Measures were averaged across both sides. Group differences were analyzed using students T-test (2-tailed) and Pearson correlations to evaluate the relationships between DTI metrics and T<sub>2</sub>.

**RESULTS-** This study is ongoing. Preliminary analyses for 3 pairs (1 *shiverer* and 1 control matched for age and gender) show remarkably similar results to the *in vivo* work of Song et al. (2002). These are illustrated in Table 1. FA measures were significantly lower ( $p \leq .05$ ) in 3 of 5 white matter regions and marginally lower ( $.05 \leq p \leq .1$ ) in the other two. Radial diffusivity measures were significantly higher ( $p \leq .05$ ) in 3 of the 5 white matter regions and marginally higher in 1 of the remaining 2. Mean axial diffusivity did not differ across groups, and no cortical DTI measures were statistically differentiated by group. There were nonsignificant trends toward higher mean T<sub>2</sub> times in *shiverer* mice in 5 of the 6 white matter ROIs. (The mean for the OT in the control set was driven by a signal high value for one control mouse.). However, in the two largest white matter ROIs (CC and IC) T<sub>2</sub> time was significantly and positively correlated with DRa in control mice (CC:  $p=.004$ , IC:  $p=.01$ ) and were correlated in IC in *shiverer* ( $p=.04$ ).

**CONCLUSIONS-** These data replicate those of Song et al. and show a relationship between developmental myelin disruption and radial diffusivity. They also extend these data by showing that diffusion tensor morphology is preserved in perfusion-fixed specimens. The data are consistent with the possibility that myelin bound water contributes to a short-TE component of the T<sub>2</sub> relaxation behavior in white matter, though the study is currently underpowered to demonstrate group differences in overall T<sub>2</sub> time in the ROIs examined. *Shiverer* may prove to be a useful model for studying myelin-based white matter abnormalities in human disease processes.

**Table1 DTI Measures and T2 Time in *shiverer* and control Mice**

REGION	CONTROL	SHIVERER
<u>Fractional Anisotropy (mean <math>\pm</math> sd)</u>		
AC*	.72 $\pm$ .05	.65 $\pm$ .01
CC**	.69 $\pm$ .03	.63 $\pm$ .01
IC*	.64 $\pm$ .02	.51 $\pm$ .09
OT***	.86 $\pm$ .05	.67 $\pm$ .04
FX**	.57 $\pm$ .05	.48 $\pm$ .02
CTX <sup>NS</sup>	.33 $\pm$ .05	.35 $\pm$ .02
<u>Axial Diffusivity (mean <math>\pm</math> sd)</u>		
AC <sup>NS</sup>	.84 $\pm$ .05	.88 $\pm$ .02
CC <sup>NS</sup>	.76 $\pm$ .08	.85 $\pm$ .03
IC <sup>NS</sup>	.73 $\pm$ .07	.77 $\pm$ .03
OT <sup>NS</sup>	.99 $\pm$ .09	.89 $\pm$ .05
FX <sup>NS</sup>	.66 $\pm$ .05	.69 $\pm$ .03
CTX <sup>NS</sup>	.75 $\pm$ .05	.72 $\pm$ .06
<u>Radial Diffusivity (mean <math>\pm</math> sd)</u>		
AC <sup>NS</sup>	.23 $\pm$ .03	.26 $\pm$ .01
CC***	.22 $\pm$ .01	.29 $\pm$ .01
IC**	.24 $\pm$ .01	.32 $\pm$ .05
OT**	.19 $\pm$ .02	.25 $\pm$ .03
FX*	.27 $\pm$ .04	.32 $\pm$ .01
CTX <sup>NS</sup>	.33 $\pm$ .05	.35 $\pm$ .02
<u>T2 time in msec (mean <math>\pm</math> sd)</u>		
AC <sup>NS</sup>	38.9 $\pm$ 5.8	39.4 $\pm$ 1.3
CC <sup>NS</sup>	38.9 $\pm$ 3.7	40.5 $\pm$ 2.0
IC <sup>NS</sup>	35.5 $\pm$ 3.1	38.3 $\pm$ 0.6
OT <sup>NS</sup>	42.6 $\pm$ 5.6	40.1 $\pm$ 1.4
FX <sup>NS</sup>	37.8 $\pm$ 2.4	40.5 $\pm$ 2.3
CTX <sup>NS</sup>	40.6 $\pm$ 0.9	34.5 $\pm$ 10.6

\*  $p \leq .10$   
 \*\*  $p \leq .05$   
 \*\*\*  $p \leq .01$