

TmDOTMA⁺: A sensitive MR thermometry probe for in vivo applications

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Introduction

Non-invasive temperature monitoring has many direct uses in medicine. MR thermometry techniques based on the chemical shift, relaxation rates and molecular diffusion rate of ¹H water signal suffer from poor thermal resolution. Zhu *et al.* [1] and we [2] independently developed a non-invasive MR thermometer based on the temperature dependence of hyperfine shifted ¹H signal of the paramagnetic lanthanide complex, TmDOTA⁺. One potential drawback of TmDOTA⁺ is low signal-to-noise ratio. In this study, we evaluate the use of lanthanide complexes of a methyl substituted analog of DOTA⁴⁻, DOTMA⁴⁻ for MR thermometry. Aime *et al.* [3] have explored the utility of Yb-DOTMA⁺ as temperature sensitive imaging probe. DOTMA⁴⁻ has 12 equivalent protons on the four methyl groups and gives three times more intense signal compared to TmDOTA⁺. In addition, the methyl proton signals have longer T_2 and narrower line-widths because of fast free rotation of the CH₃ groups and reduced through-bond paramagnetic contact interaction.

Experimental

TmDOTMA⁺ were synthesized from Ln₂O₃ (Ln = Pr, Yb, Tb, Dy and Tm) and Na₄DOTMA. ¹H spectra of the LnDOTMA⁺ complexes were acquired in the temperature range 22 to 55°C using a Varian 9.4 T 89 mm vertical bore MR system. ¹H spin-lattice (T_1) and spin-spin (T_2) relaxation times of the methyl resonances from the five LnDOTMA⁺ complexes were measured at 37°C. To determine the effects of pH and Ca²⁺, on the chemical shift of the methyl resonance of the TmDOTMA⁺, the experiments were conducted at 37°C at five different pH values (3 to 11) and five different Ca²⁺ concentrations (0 to 3 mM). *In vivo* temperature measurements were performed on subcutaneously (sc) implanted RIF-1 tumor in C3H/HeN mice using a 1 cm diameter surface coil placed over the tumor. 1-2 mmole of TmDOTMA⁺ per kilogram body weight was injected through a tail vein. Animal core body temperature was monitored with a rectal fiber-optic temperature probe. The animal temperature was manipulated over temperature ranging from 35 to 40°C by blowing warm air into the magnet bore.

Results and Discussion

Figure 1 shows the structure and ¹H MR spectra of TmDOTA⁺ and TmDOTMA⁺ showing H¹ and H⁶ or methyl proton signals. The SNR advantage with TmDOTMA⁺ is clearly apparent in the spectra. Table 1 shows the chemical shifts, temperature coefficients of chemical shift (C_T), line-widths and relaxation times for the methyl resonance from the five complexes. Tb(III), Dy(III), and Tm(III) complexes of DOTMA⁴⁻ show two resonances because of the presence of two conformational isomers. The relative amount of the minor isomer of TmDOTMA⁺ is < 4%. The C_T value is the largest for the methyl signal from TmDOTMA⁺. The ratio of temperature coefficient and resonance full width at half height ($|C_T|/\text{FWHH}$) is also largest for TmDOTMA⁺, therefore, this complex was further evaluated. The methyl proton chemical shifts of TmDOTMA⁺ are independent of the pH, Ca²⁺ concentration or presence of any blood plasma. The proton T_1 and T_2 values for the methyl resonance of TmDOTMA⁺ at 37°C and 9.4 T are 5.3 and 4.1 ms, respectively. These values are approximately two times more compared to the H⁶ signal from TmDOTA⁺. These data clearly show the advantages of TmDOTMA⁺ over TmDOTA⁺. Figure 2 shows representative *in vivo* ¹H spectra of TmDOTMA⁺ from a sc-implanted RIF-1 tumor. The tumor temperature was always lower than the core body temperature. This demonstrates that TmDOTMA⁺ allows robust measurement of temperature in sc implanted tumors and other tissue in intact animals.

Conclusion

The major advantages of TmDOTMA⁺ for MR studies include 1) ~60 times more sensitivity to temperature than water and 15 times more than YbDOTMA⁺; 2) more intense signal and longer T_2 compared to TmDOTA⁺ [1,2]; and 3) insensitivity to changes in concentration, pH, [Ca²⁺] and presence of other ions and macromolecules. These properties should make TmDOTMA⁺ useful for MR thermometry in a wide range of applications.

References

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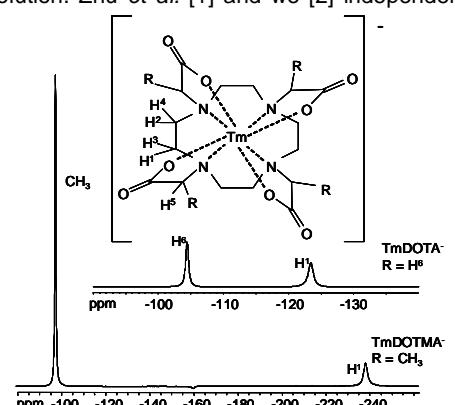


Fig. 1: ¹H MR Spectra of TmDOTA⁺ and TmDOTMA⁺ showing H¹ and H⁶ or methyl proton signals. The H¹ resonances from both the complexes is set to the same signal intensity demonstrating the SNR advantage with the methyl resonance from TmDOTMA⁺.

Table 1: ¹H chemical shifts, temperature coefficients (C_T), full-width at half-height (FWHH), $|C_T|/\text{FWHH}$, T_1 and T_2 for ¹H methyl signal from Pr(III), Yb(III), Tb(III), Dy(III) and Tm(III) complexes DOTMA⁴⁻.

Lanthanide complex	^a Shift ppm	^b C_T ppm/°C	^c FWHH ppm	^c $ C_T /\text{FWHH}$	^d T_1 ms	^d T_2 ms
PrDOTMA	6.80	-0.014	0.065	0.210	90	17
YbDOTMA	-13.9	0.058	0.063	0.933	59	29
^e TbDOTMA	63.3	-0.269	0.90	0.300	3.0	1.3
	57.5	-0.130	0.48	0.273		
^e DyDOTMA	73.3	-0.233	0.59	0.395	2.2	1.7
	78.0	-0.325	0.93	0.351		
TmDOTMA	-99.6	0.586	0.43	1.362	5.3	4.1
	-67.1	0.184	0.39	0.473		

^a at 35 °C;

^b Slope of shift vs temperature data from 22-55 °C.

^c Absolute value of the ratio of temperature coefficient of chemical shift (C_T in ppm/ °C) and the FWHH (full width at half height) at 35 °C (in ppm).

^d Represents the existence of the isomers for each of the methyl group.

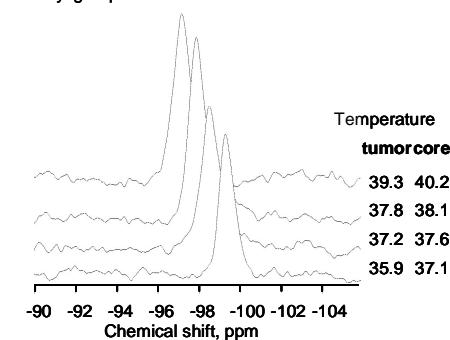


Fig. 2: Representative *in vivo* ¹H spectra from methyl resonance of TmDOTMA⁺ from sc-implanted RIF-1 tumor. The tumor temperature was calculated from the chemical shift of the TmDOTMA⁺ methyl proton signal with respect to the water proton signal set to 4.7 ppm. The core temperature was measured using a fibre-optic rectal probe.