Study of 17O NMR Sensitivity and Relaxation Times of Cerebral Water in Human at 7 Tesla

X. Zhu¹, X. Zhang¹, W. Chen¹

¹Center for Magnetic Resonance Research, University of Minnesota, Minneapolis, Minnesota, United States

Synopsis The cerebral metabolic rate of oxygen (CMRO₂) is an important physiological parameter for studying brain function and diseases. We demonstrated previously the possibility of using high-field ^{17}O MR approach for mapping metabolic $H_2^{17}O$ and imaging CMRO₂ in rat brain. Recently, we have studied ^{17}O relaxation times of the natural abundance $H_2^{17}O$ in human brain (T_1 =5.46±0.09ms, T_2 =4.32±0.24ms and T_2 *=2.40±0.07ms), and sensitivity of 3D ^{17}O MRS image at 7 Tesla. Our results indicate that the excellent sensitivity of ^{17}O MRS image obtained at ultra-high field could be feasible for achieving 3D imaging of CMRO₂ in the human brain. **Introduction**

In oxidative tissues such as the brain, oxygen is primarily utilized through oxidative phosphorylation in the mitochondrial respiratory chain according to the reaction: $4H^+ + 4e + O_2 \rightarrow 2H_2O, \qquad [1]$

and this process is tightly associated with the generation of high-energy phosphate compounds. Questions that involve the cerebral metabolic rate of oxygen consumption (CMRO₂) are encountered frequently in biomedical research when considering either normal tissue function or abnormalities induced by brain diseases. It is, therefore, important to have methods for measuring and imaging CMRO₂ accurately and noninvasively. The simplest NMR approach to determine CMRO₂ is to detect the metabolic H₂¹⁷O directly using ¹⁷O NMR approach during inhalation of ¹⁷O labeled oxygen gas (1-3). However, the spatial resolution of ¹⁷O NMR is limited by the low gyromagnetic ratio of ¹⁷O nucleus and consequently its relatively poor sensitivity, and the extremely low H₂¹⁷O concentration metabolized from inhaled ¹⁷O₂ gas. Therefore, the key factor for determining CMRO₂ using ¹⁷O approach is the sensitivity of ¹⁷O NMR signal acquired within a unit time, which is given by

 $SNR \propto B_o^{\beta} (T_2^*/T_1)^{1/2},$ [2]

where B_0 is magnetic field strength, T_1 is longitudinal relaxation time and T_2 * is apparent transverse relaxation time.

Recently, our results showed that the relaxation times of ¹⁷O water are field *independent* presumably due to the dominance of quadrupolar relaxation, and the ¹⁷O NMR sensitivity increased approximately *four-fold* at 9.4T compared to at 4.7T in the rat brain (4). With this significant sensitivity gain, we have successfully applied the 3D ¹⁷O MRS imaging method to determine CMRO₂ in the anesthetized rat brain at 9.4 Tesla during a two-minute ¹⁷O₂ inhalation (5). In this study, we have extended our efforts toward the human brain study at 7 Tesla for determining the T₁, T₂ and T₂* relaxation times of the natural abundance water, as well as ¹⁷O NMR sensitivity in the 3D ¹⁷O MRS images with a voxel size of 6.6 ml.

Methods

All experiments were conducted on a 90 cm bore 7 Tesla Magnex magnet with a Varian INOVA console. A multinuclear surface-coil probe consisting of a 5cm-diameter ^{17}O surface coil (40 MHz) and a larger quadrature ^{1}H coil (300 MHz) was used for detecting NMR signals from the human visual cortex. Non-localized spin-echo (θ -TE/2-2 θ -TE/2-acquisition) with phase cycling and inversion recovery (180° -TI- θ -acquisition) pulse sequences were applied to determine T_2 and T_1 values of natural abundance $H_2^{17}\text{O}$, respectively, where θ is the flip angle of a hard square pulse (200 µs). A total of 12 TE and 8 TI values were used to calculate T_2 and T_1 values. The T_2 * values were calculated by T_2 * = $1/(\pi \Delta v_{1/2})$, where $\Delta v_{1/2}$ is the line width of T_2 0 resonance peak. Other NMR parameters were: TR = 33 ms (> 5 T₁); scan number = 128-256; spectral width = 30 kHz. The spatial localization of T_2 0 MRS was achieved by using the 3D Fourier series window (FSW) MRS imaging technique (6). The acquisition time for each 3D T_2 0-MRS image was 8.5 seconds (total scan number=254; spectral width=30 kHz; FOV=9.4×9.4×7.5 cm³, 7×7×5 phase encodes). The image voxel size was 6.6 ml. The FIDs were zero-filled and a 100-Hz line broadening was used before Fourier transformation for SNR enhancement. The averaged RF power used in this study was below the FDA limit of SAR. Four human subjects participated this study. The institutional review board at University of Minnesota approved all study procedures.

Results

The ^{17}O relaxation times in the human visual cortex were found to be: $T_1 = 5.46 \pm 0.09$ ms, $T_2 = 4.32 \pm 0.24$ and $T_2 * = 2.40 \pm 0.07$ ms (n=4) at 7 Tesla. The averaged linewidth of ^{17}O resonance peak of cerebral water is 133 ± 4 Hz (n=4). The 3D ^{17}O MRS imaging technique was applied to measure the natural abundance $H_2^{17}O$ distribution in the human brain with high temporal resolution of 8.5 seconds and 6.6 ml voxel size. Figure 1a demonstrates results obtained from a representative subject showing one 2D coronal ^{17}O chemical shift image (CSI) of natural abundance $H_2^{17}O$ extracted from 3D ^{17}O image dataset acquired using the ^{17}O coil. The CSI slice was located in the visual cortex. The spatial distribution of ^{17}O NMR signal intensity is not uniform because of the inhomogeneous B_1 field of the ^{17}O surface coil used. Figure 1b displays a representative ^{17}O spectrum from one voxel showing the resonance peak of natural abundance $H_2^{17}O$ in the human visual cortex. Excellent SNR was achieved with a relatively short acquisition time at 7 Tesla, especially for the central voxels where SNR was optimized by the B_1 profile of the ^{17}O coil (SNR \approx 60:1) as demonstrated in Fig. 1b.

Discussion and Conclusions

The 17 O relaxation times in the human brain at 7 Tesla are similar to that of rat brain at 9.4 Tesla ($T_1 = 4.84 \pm 0.18$ ms, $T_2 = 3.03 \pm 0.09$ ms, $T_2 = 1.80 \pm 0.06$ ms) (4), however, they are significantly higher than the values determined in the rat brain. These increases may reflect the differences between different species. The relatively large ratio of ($T_2 * / T_1)^{1/2}$ (=0.66) of human brain at 7 Tesla in comparison with the ratio (=0.61) in the rat brain at 9.4 Tesla will gain ~8% SNR according to Eq. [2]. Although 17 O nucleus has a relatively low gyromagnetic constant (only

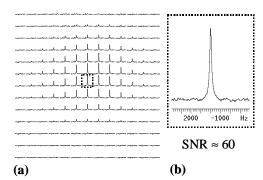


Fig. 1 (a) 3D 17 O CSI images (displayed in 2D) of natural abundance H_2^{17} O in the human brain at 7T, and (b) central voxel 17 O spectrum.

14% versus that of the proton nucleus), excellent 17 O spectra of cerebral natural abundance water were obtained in the human brain by taking the advantage of ultrahigh magnetic field and optimizations of NMR acquisition parameters (e.g., fast acquisition due to short T_1). This sensitivity made it possible to obtain a 17 O-MRS image of natural abundance cerebral H_2 17 O with a relatively high temporal resolution (8.5 s) in the human brain. Note that although the current voxel size used for the human brain is much larger than that of the rat brain, it is comparable to that provided by most conventional PET scanners for imaging CMRO₂ (7). The 17 O sensitivity and the temporal resolution of image acquisition in the human 3D 17 O MRS image is slightly better than that used in the rat brain study. Furthermore, the dynamic concentration change of metabolic H_2 17 O during 17 O₂ inhalation is expected to be similar between the awaked human brain and the rat brain anesthetized with α -chloralose because of their similar values of both CMRO₂ (2.2 µmol/g/min in rat versus 1.7 µmol/g/min in human) and cerebral blood flow (0.53 ml/g/min in rat versus 0.54 ml/g/min in human) (5, 7). In summary, our results indicate the feasiblity for achieving 3D imaging of CMRO₂ in the human brain at 7 Tesla. Realization and utilization of this feasibility would have a great impact on studying the central role of oxidative metabolism in brain activation and pathological diseases.

Acknowledgments: NIH grants NS41262, NS38070, NS39043, EB00329, P41 RR08079, the W.M. Keck Foundation and the MIND institute.

References: [1] Mateescu GD et al. Proceedings of SMRM, 1989. p 659. [2] Arai T et al. Biochem Biophys Res Commun 1991; 179:954-61. [3] Pekar J et al. Magn Reson Med 1991; 21:313-9. [4] Zhu XH et al. Magn Reson Med 2001; 45:543-9. [5] Zhu XH et al. Proc Natl Acad Sci USA 2002; 99:13194-13199. [6] Hendrich K et al. J Magn Reson 1994; 105:225-232. [7] Fox PT et al. Science 1988; 241:462-464.