Observation of label accumulation at Glutamate/Glutamine $C_{1,2,3,4}$ and HCO_3 in human brain after intravenous 1-13C labeled Glucose infusion

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Introduction:

Glucose (Glc) is the major respiratory fuel of the human brain generating 36 ATP's per molecule if completely oxidized to $CO_2 + H_2O$. Decarboxylation steps include pyruvate dehydrogenase, isocitratedehydrogenase, and oxoglutarate dehydrogenase. Proof of complete oxidation comes from the detection of CO₂ (or HCO₃⁻) formation. Several groups demonstrated that once intravenously (i.v.) infused 1-13C Glc passes the blood-brain barrier label accumulation in glutamate (Glu), glutamine (Gln), y-amino butyric acid (GABA), lactate (Lac), and aspartate (Asp) can be detected (1-7). These experiments in humans did not allow the observation of signals resonating at > 150 ppm and enrichment of HCO₃, Glu₁ and Gln₁, as shown in tissue slices and extracts of animal brain (8,9), was not reported. Aims: (i) Establish assignments and enrichment patterns in human brain after Glc infusion for resonances at chemical shifts > 150 ppm. (ii) Demonstrate CO₂ production from glucose oxidation in vivo.

Material and Methods:

Two adult controls (male 27 years, fed, female 29 years, fasted) were studied with natural abundance and with ¹³C MRS after i.v. glucose infusion using a protocol modified from that developed by DeFronzo et al. (10) by omitting somatostatin. Experiments were carried out on a GE, Signa 1.5 T system using a coil design as described previously (11,12). A baseline spectrum and spectra during infusion were acquired from the occipital brain region with a FID sequence, TR = 1s, 1024 pts, 4 kHz excitation bandwidth.

Results:

As in earlier studies, labeling of $Glu_{2,3,4}$, $Gln_{2,3,4}$, $Asp_{2,3}$, and $NAA_{2,3}$ was observed (Fig. 1). In the region > 150 ppm the chemical shifts of the ^{13}C enriched resonances at 175.3 and 174.8 ppm, are consistent with Glu_1 and Gln_1 , the resonance at 160.9 ppm is consistent with carbonate (HCO_3^-) (Fig. 2). ^{13}C enrichment of these resonances was observed in both subjects after ≈ 50 minutes.

Discussion:

These findings represent an extension of previous studies (4-6) where enrichment of $Glu_1,\ Gln_1,\ and\ HCO_3^-$ was not reported. Our data suggest that proton decoupled ^{13}C MRS allows a more detailed definition of the fates of $1^{-13}C$ glucose carbons, including the accumulation in $Glu_1,\ Gln_1,\ and\ HCO_3^-,\ than previously available. This potentially will provide a more precise simultaneous determination of the TCA-cycle flux rate, glutamine synthesis rates, malate-aspartate shuttle exchange rate, <math display="inline">\alpha$ -ketoglutarate/glutamate exchange rates, now including substrate decarboxylation and total glucose oxidation in normal and diseased human brain.

References

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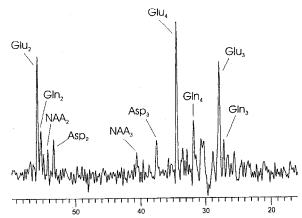


Fig. 1: A difference spectrum, generated by subtracting the baseline scan from the spectrum acquired between 75-145 min after infusion start from the male subject, shows label accumulation in various amino acid resonances between 20-60 ppm chemical shift.

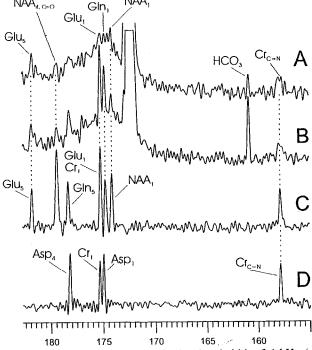


Fig. 2: Due to the large excitation bandwidth of 4 kHz (\approx 200 ppm) signal from metabolites resonating at > 150 ppm can be observed simultaneously. Shown is the baseline spectrum (A), acquisition time 25 min, and the spectrum acquired between 140 - 180 min (B), aligned with spectra from model solutions of Glu, Gln, NAA, and Cr (C), and Asp and Cr (D) from the female volunteer. A \approx 4% enrichment of HCO3 can be observed. Two resonances consistent with Glu₁ and Gln₁ are also clearly enriched.

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