Articular cartilage injuries are common types of injuries encountered in orthopaedic practice. Several therapeutic strategies have been developed to repair damaged articular cartilage and produce a durable repair. One of the components of healthy cartilage, glycosaminoglycans (GAG), is responsible for the biomechanical properties of articular cartilage [1], and focal loss of GAG represents the earliest stages of cartilage degeneration [2]. GAG specific MR techniques are delayed gadolinium-enhanced MRI of cartilage (dGEMRIC [3]) and sodium imaging [4]. Recently, a new GAG-specific sequence based on chemical exchange was reported [5]. The aim of this study was to investigate a newly developed method, which enables 3-dimensional gagCEST imaging in clinically relevant scan times at 7 Tesla. Results of gagCEST imaging in patients after cartilage repair surgery were compared to results obtained with another glycosaminoglycan-specific technique, namely sodium imaging, as a reference method.

Materials & Methods:
The study comprised 12 patients who underwent cartilage repair surgery. All examinations were performed in accordance and with approval of the local ethics committee. Experiments were performed on a 7 T MRI system (Siemens Healthcare, Erlangen, Germany). For 1H imaging, a new 28-channel Tx/Rx knee coil (QED, Mayfield Village, OH, USA), and for 23Na imaging a circularly polarized knee coil (Stark Contrast, Erlangen, Germany) were employed. For gagCEST imaging, a series of Gaussian-shaped RF pulses followed by gradients in x-, y- and z-direction was applied to saturate proton resonances at variable offset frequencies to each spectral side of bulk water. After RF pre-saturation, image readout was performed with a 3-dimensional RF-spoiled gradient echo (GRE) sequence (TE = 3.2 ms, TR = 7.3 ms). The FoV was 146 x 180 x 84 mm³ with a matrix size of 208 x 256 x 28. Total acquisition time for one CEST measurement was 14:34 min. Sodium imaging was performed with a modified 3-dimensional GRE sequence (TE = 3.77 ms, TR = 10 ms) [6]. The acquisition matrix was 64 x 128 x 36 with a FoV of 199 x 199 x 108 mm³, and the measurement time was 30:45 min.

To compensate for movement of the knee during the course of a measurement, gagCEST datasets were registered using a non-rigid approach. Z-spectra from images were fitted with a cubic spline method and corrected for B₀ inhomogeneities on a pixel-by-pixel basis. The asymmetry of the magnetization transfer rate (MTR) as determined by $\text{MTR}_{\text{gagCEST}}(\delta) = \text{MTR}(\delta) - \text{MTR}(0)$ was summed for offsets from 0 – 1.3 ppm in each pixel and used as gagCEST signal intensity. Sodium SNR values of ROIs were used for data analysis of sodium images to ensure comparability between subjects. Sodium and gagCEST images were examined with regions-of-interest (ROI) analysis. Sodium SNR values and gagCEST signal intensities were measured from a region covering the cartilage repair tissue and an equal region size region in normal native cartilage. To assure comparability between ROI placement in the two imaging modalities similar sized ROIs were drawn in the cartilage repair tissue and the native cartilage, in the same positions on identical slices in a side-by-side evaluation. Ratios of ROI values obtained in healthy cartilage compared to repair tissue were determined and the Pearson correlation coefficient was calculated to evaluate associations between gagCEST and sodium imaging.

Results:
The average asymmetry in gagCEST z-spectra summed over all offsets from 0 to 1.3 ppm was significantly higher ($p < 0.004$) in native cartilage (8.01 ± 2.79 % (mean ± SD)) than in repair tissue (4.65 ± 2.23 %) (Fig. 1). This is in good agreement with sodium imaging, where healthy cartilage (16.53 ± 3.66) yielded significantly higher ($p < 0.004$) higher SNR compared to repair tissue (12.36 ± 2.46) (Fig. 2). Furthermore, a strong correlation ($R = 0.885, p = 0.0001$) was found between ratios of native cartilage to repair tissue obtained with gagCEST, and sodium imaging (Fig. 3). The mean dimensionless ratios between native cartilage and repair tissue were 1.39 ± 0.30 for gagCEST, and 1.34 ± 0.21 for sodium MRI. Delineation of repair tissue from native cartilage was equally well possible with both techniques employed in this study. Localizations of signal reductions as found in sodium images showed good agreement with gagCEST images (Fig. 4).

Discussion and Conclusion:
The high correlation between results obtained with the new gagCEST method, and ²³Na imaging, respectively, demonstrate the GAG specificity and reliability of the developed gagCEST imaging method. Both techniques suggest that cartilage repair tissue has lower GAG content than native tissue. This study used the beneficial properties that high magnetic field offers for both techniques. Since SAR restrictions, patient compliance, and availability of 7 T whole-body MR scanners would limit broader use of the techniques for clinical routine, a transition of the gagCEST method to 3 T is currently under investigation.

References: