Sensitive MRI markers for systemic amyloidosis: amide proton transfer and equilibrium contrast

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Introduction: Systemic amyloidosis is a fatal disorder characterised by the extracellular deposition of insoluble abnormal protein fibrils. Amyloidosis remains a clinical challenge due to difficulties in diagnosis and treatment [1]. To address these challenges, our aim was to develop sensitive MRI markers for the assessment of amyloid accumulation, which could be used to investigate novel treatments in mouse models and provide diagnostically relevant information in the clinical setting.

In this study, two novel MRI techniques were applied to amyloidosis: equilibrium contrast MRI (EQ-MR) and amide proton transfer (APT) imaging. EQ-MR relates the volume of distribution (Vd) of gadolinium (Gd), a contrast agent that accumulates in the extracellular matrix, to the expansion of interstitial space in extracellular pathologies such as amyloidosis [2]. APT imaging describes the specific application of chemical exchange saturation transfer (CEST) imaging for detection of amyloidosis [3], used here for the detection of protein accumulation associated with amyloidosis. Additionally, cine MRI was applied to analyse cardiac function (ejection fraction, stroke volume, left-ventricle mass, etc) [4].

Methods: Ten control and eleven amyloidotic mice, with inducible transgenic over-expression of serum amyloid A protein (SAA) [5], were studied. Imaging was performed on a Varian 9.4T scanner (Varian Inc. Palo Alto, CA, USA) with 35mm coil (RAPID Biomed, Rimpar, Germany).

EQ-MR: A mid-ventricle short-axis slice through the heart, which included a section of liver, was used for EQ-MR. A primed-infusion of Magnevist (Schering, Berlin) was used to generate a blood - tissue equilibrium of [Gd]. The dose and timing protocol was optimised empirically such that equilibrium was reached as quickly as possible. A bolus of 0.6mmol/kg was followed 10 minutes later by 0.005mmol/kg/minute infusion of Magnevist. An ECG-gated Look-Locker technique [6] was used to measure the T1 pre- and post-contrast (TE/TR = 1.18ms/13.5s/RR-interval, flip-8°, in-plane resolution=200μm, slice thickness=1.5mm). A 3-parameter fit was used to generate T1 values for regions-of-interest (ROIs) in the myocardium, liver and left ventricle blood pool. The Vd of Gd in the myocardium and liver was calculated using:

\[ V_d = \frac{\Delta S_{\text{meas}}}{\Delta S_{\text{blood}}} \times S_0 \]

where \( R_1 = 1/T_1 \) and Hct is the group mean hematoctrit, measured from tail-vein blood samples.

APT: APT imaging was applied in the kidney. Axial gradient-echo images (TE/TR = 2.5ms/6s, flip angle = 20°, in-plane resolution = 200μm, slice thickness = 1mm) of the kidney were acquired following RF preparation pulses (100 Gaussian pulses) with top-up saturation pulses between lines of k-space. Preparation pulses of frequencies -3500Hz to +3500Hz in 50Hz steps, corresponding to a range of ±0.75ppm, were used to generate a CEST-spectrum. The magnetisation transfer asymmetry (MTA) was calculated using:

\[ MTA\% = \frac{S_m - S_0}{S_0} \times 100\% \]

where S is the signal intensity of the images, with preparation pulses at a frequency indicated by the subscript and \( S_0 \) is a reference measurement at a frequency of 8000Hz. MTA was calculated pixel-wise then averaged for ROIs in the kidney cortex, medulla and pelvis. The %APT was calculated from the mean MTA of the amide proton peak (2.5-4.5ppm) [3]. All image analysis for the EQ-CMR and APT techniques was performed in MATLAB.

Cine MRI: Global cardiac function was calculated from double gated spoiled gradient echo cine images (TE/TR = 1.2/4.5-5ms, cine frames= 20, in-plane resolution=200μm, slice thickness = 1mm, 10 short-axis slices) using the freely available software Segment (http://segment.heiberg.se/).

Results: We confirmed by histology that the amyloidotic animals had major amyloid deposits in the liver and minor deposits in their kidneys and heart, as is characteristic of this model [5]. Analysis of cardiac functional parameters calculated from cine images showed no significant difference between the groups. However, using EQ-MR, the amyloidotic group shows a significantly increased Vd of Gd compared to the control group in both organs (Figure 1). The Vd of the control group was 15.4±1.5% (myocardium) and 15.4±2.7% (liver) and of the amyloidotic group 19.8±4.1% (myocardium) and 23.6±4.6% (liver) (mean ± s.d.). Figure 2 presents MTA spectra of the two groups for the a) cortex, b) medulla c) pelvis and d) mean of all regions. The amyloidotic group shows a significant increase in %APT in the kidney cortex (+2.2±2.5% (control) vs. 0.8±1.3% (amyloidotic), but there was no significant difference in the other regions. Also, a significant difference was observed when the mean of the regions is taken (-0.7±3.1% (control) vs. 1.5±1.5% (amyloidotic)) (mean ± s.d.).

Discussion: In this study, both EQ-MR and APT techniques were able to detect minor amyloid deposits. The EQ-MR procedure has been optimised in the mouse and applied to image the liver and heart. The results of this study show that EQ-MR techniques can detect low levels of amyloid in the heart before impaired cardiac morphology or function can be observed by cine imaging. This approach has the potential to become a sensitive diagnostic tool.

In conclusion, we present two novel MRI techniques for early detection of amyloidosis. We found that both EQ-MR and APT were useful in detecting minor amyloid deposition, and EQ-MR appeared to be more sensitive than examination of cardiac function. Our results demonstrate that these MRI techniques should be useful for early diagnosis of systemic amyloidosis as well as enable serial quantitative monitoring of response to novel therapies aimed at elimination of amyloid deposits in preclinical studies [7].